

## SHORT INSTRUCTION

# FastQ<sup>®</sup> B\*27 direct

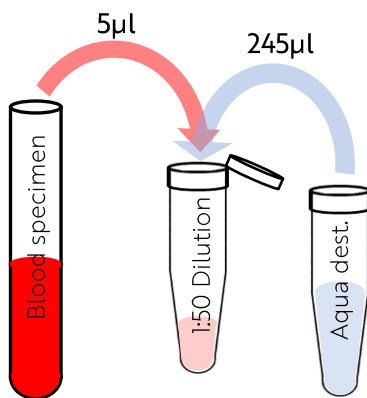
REF 728201



### PCR PROGRAM

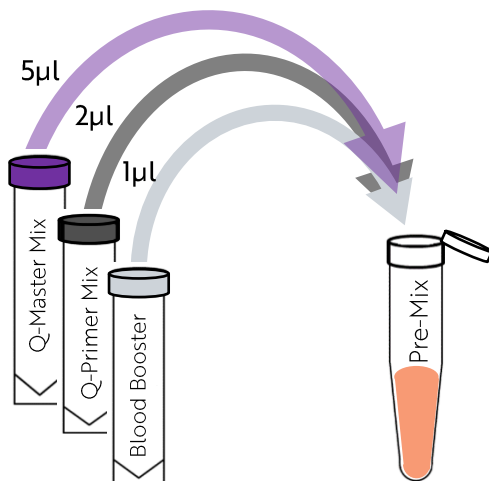
Step	Time [sec]	Temperature [°C]	Ramp Rate [°C/sec]	Cycles
Initial Activation	120	96	2.5	1
Denaturation	5	98	2.5	18
Annealing + Extension	25	64	2.2	
Denaturation	5	98	2.5	42
Annealing + Extension	25 + Plate Read	64	2.2*	
Cooling	120	37	2.2	1


\* use the default ramp rate of the CFX96 Touch & CFX Opus 96 Real-Time PCR Detection System

### WORKFLOW



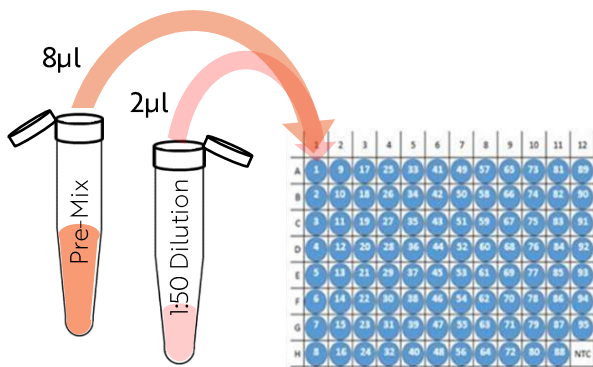
1. Sample Dilution (1:50)	
	Mix specimen
245 µl	Add aqua dest.
5 µl	Pipet blood into 245 µl aqua dest.
	Mix sample dilution



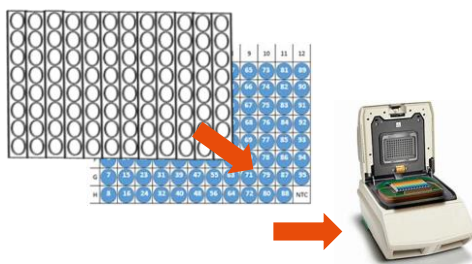
2. Pre-Mix Preparation	
5 µl	Q Mastermix
2 µl	Q Primermix B27-d
1 µl	Blood Booster
<b>Total 8 µl</b>	<b>Pre-Mix</b>
	Mix Pre-Mix thoroughly

**Pre-Mix Setup Table**

No. of tests (n)	Q Primermix [µl]	Q Mastermix [µl]	Blood Booster [µl]	Pre-Mix Volume [µl]
1	2	5	1	8
8	20	50	10	80
16	36	90	18	144
24	56	140	28	228
32	72	180	36	292
40	88	220	44	356
48	104	260	52	420
56	120	300	60	484
64	140	350	70	566
72	156	390	78	630
80	172	430	86	694
96	208	520	104	840



3. Pre-Mix and Sample Distribution	
	Mix Pre-Mix
8 µl	Pre-Mix
	Mix Sample Dilution
2 µl	Sample (Dilution 1:50)
<b>Total 10 µl</b>	<b>Reaction Volume</b>



4. Start RT-PCR	
Close plate with caps or foil	
Place plate into RT-cycler and start the run	

**INTERPRETATION**

Specificity	Fluorophore	Baseline Threshold	Quantification cycle (Cq)
B*27	Cal Fluor Orange 560	Auto	≤ 30
Internal Amplification Control (IAC)	FAM	Auto	≤ 20