

## SHORT INSTRUCTION

# FastQ<sup>®</sup> B\*27

**REF** 728208

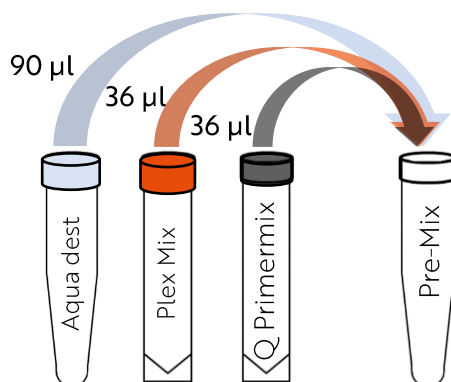
### CREATE RUN ID IN THE PlexTyper<sup>®</sup> SOFTWARE (OPTIONAL)



Enter the sample information in the software and save the run to create the **RUN ID** (PT1, PT2, PT3....) that is used to track the test.

### PCR PROGRAM

Step	Time	Temperature [°C]	Ramp Rate [°C/sec]	Cycles
Initial Activation	120 sec	96	2.5	1
Denaturation	5 sec	98	2.5	13
Annealing + Extension	25 sec	68	2.2	
Denaturation	5 sec	98	2.5	37
Annealing + Extension	25 sec + Plate read	68	-	

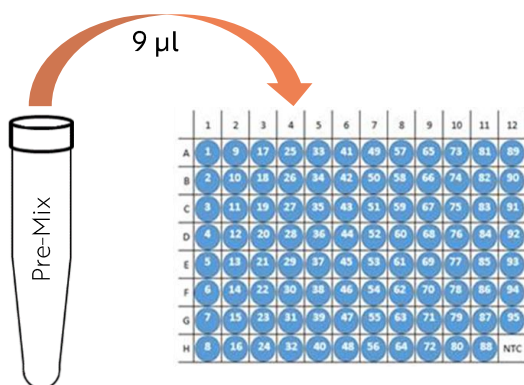
### WORKFLOW



1. Preparation (e.g. 16 reactions)	
	Mix specimen (short and careful)
90 µl	Add aqua dest.
36 µl	Q Primermix
36 µl	Plex Mix
	Mix Pre-Mix
DNA	Dilute the DNA sample → 10-20 ng/µl

### Pre-Mix Setup Table

No. of Tests (n)	Q Primermix [µl]	Plex Mix [µl]	Aqua dest. [µl]	Pre-Mix Volume [µl]
1	2	2	5	9
8	20	20	50	90
16	36	36	90	162
24	54	54	135	243
32	72	72	180	324
40	88	88	220	396
48	106	106	265	477
56	122	122	305	549
64	140	140	350	630
72	156	156	390	702
80	172	172	430	774
96	206	206	515	927



2. Dispense the Pre-Mix
Pipette <b>9 µl of Pre-Mix</b> into each well
Please take care that no liquid gets out of the well

3. Test setup
Pipette <b>1 µl of each DNA-Sample</b> into the correct well
Please take care to change the tips

4. Start real time PCR run
Close plate with caps or foil
Centrifuge the plate or knock carefully on the lab bench to avoid bubbles
Place plate into RT-cycler and start the run

### INTERPRETATION

Specificity	Fluorophore	Baseline Threshold	Quantification cycle (Cq)	Wavelength [nm]
B*27	CAL Fluor Orange 560 (HEX)	Auto	< 25	Excitation: 538 Emission: 559
Internal Amplification Control (IAC)	FAM	Auto	< 20	Excitation: 495 Emission: 520